

Short course on Methods and Tool for Genomic Predictions and GWAS in Breeding Programs

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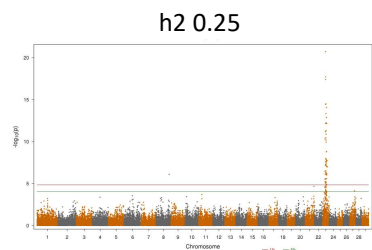


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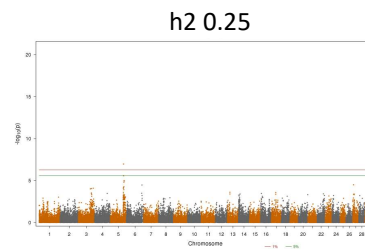
GWAS Applications in Livestock



- Understanding the genetic architecture of traits
- Uncovering causative mutations affecting economically important traits
- Improving accuracy of Genomic Selection



Major gene + polygene



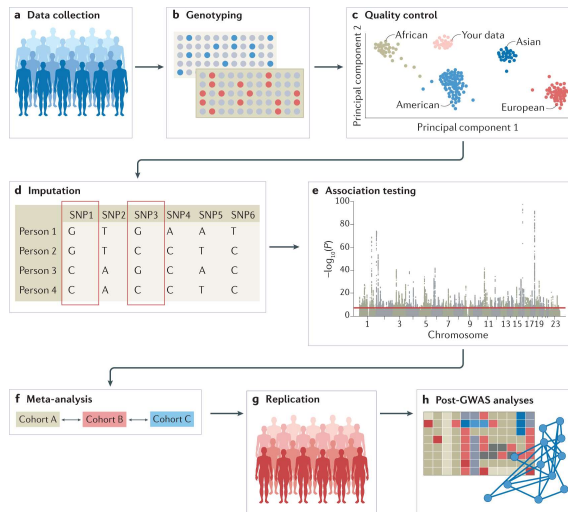
More polygenic

Results from the first 700 Ontario commercial Holsteins tested as part of the 5000 cow project – Dr. Mallard

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Steps for Conducting GWAS



Uffelmann, E., Huang, Q.Q., Munung, N.S. et al. Genome-wide association studies. Nat Rev Methods Primers 1, 59 (2021).

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Methods



- SNP by SNP GWAS
- SNP by SNP GWAS when fitting G matrix
- GBLUP/RBLUP/ssGBLUP
- Bayesian method

+Haplotype models

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Methods



- Family-based GWAS
- Random samples (Mostly common variants)
- Selective genotyping (Mostly rare variants)
- Isolated population
 - Rare variants may be observed in higher frequencies

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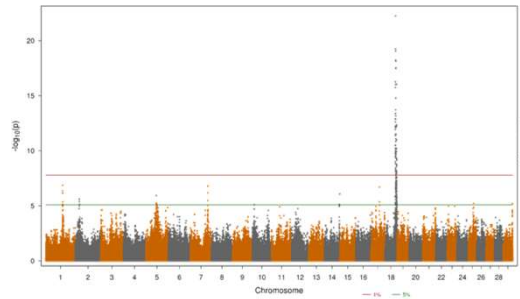
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Points to Consider When Performing GWAS

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Genome-Wide Association Studies (GWAS)



- GWAS identifies SNPs which are associated with a trait. Most of the time it cannot specify causal genes/mutations.
- Missing h^2 : Genetic variation calculated from GWAS does not completely explain the heritability of quantitative traits.

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Some Challenges



- Sample size (for mapping rare variants)
- Extent of LD and fine mapping
- Population stratification & structure
- Multiple testing (association by chance)
- Under-representation of rare variants due to ascertainment bias
- GWAS on EBV or de-regressed EBV

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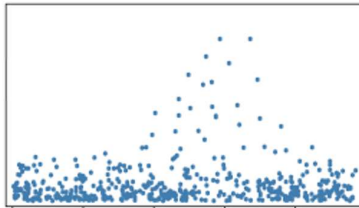
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Extent of LD and Fine Mapping

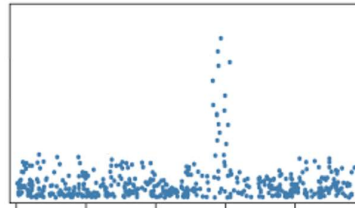


When LD is extended over long distances, fine mapping is challenging

High extent of LD



Low extent of LD

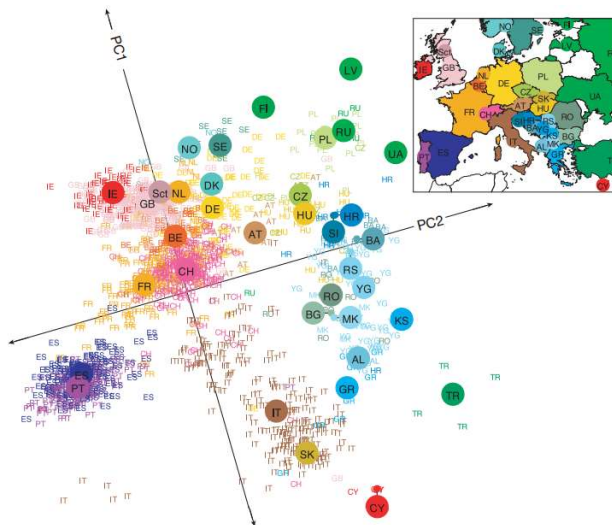


GWAS from different breeds may provide additional info to locate the causative mutation

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Population Stratification & Structure



Ref: Genes mirror geography within Europe. John Novembre et al. Nature 2008.

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Population Stratification & Structure



- Unusual allele frequency differences between subpopulations
- Phenotypes correlated with locations cause spurious associations
- Family structure or cryptic relatedness also results in spurious associations
- Systematic ancestry differences between subgroups and also admixture

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Population Stratification & Structure



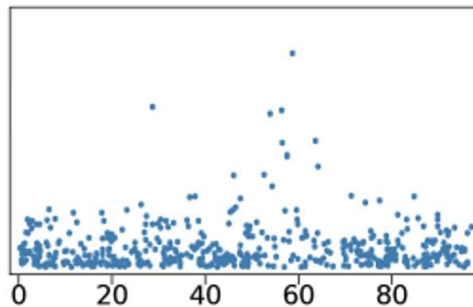
Japan

Good in Baseball

Not as good
in Baseball



Great Britain



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Population Stratification & Structure



Solution:

- Structured association (clustering)
- Fitting principal components as covariates in the model
- Fitting genomic relationship matrix in the model (Genomic control)
- GBLUP/RBLUP/ssGBLUP

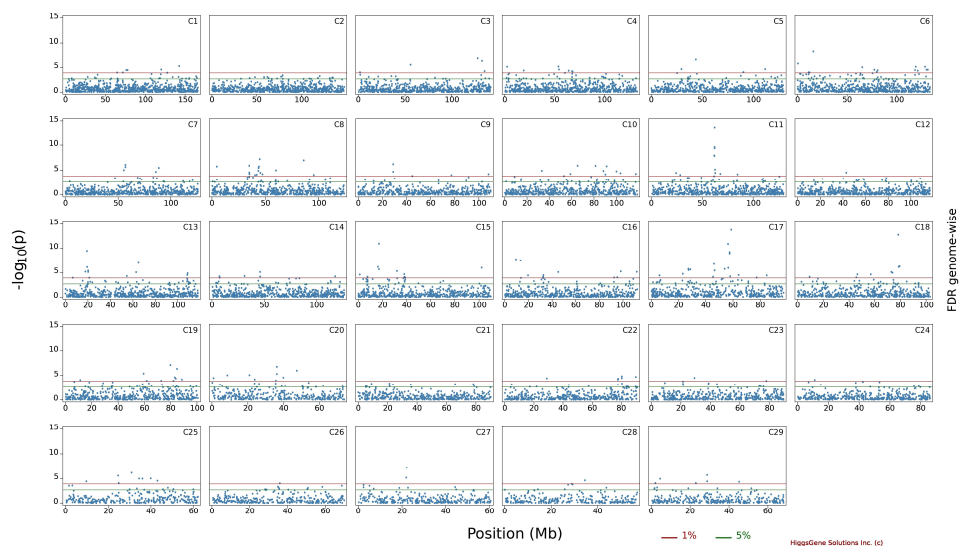
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Population Stratification & Structure



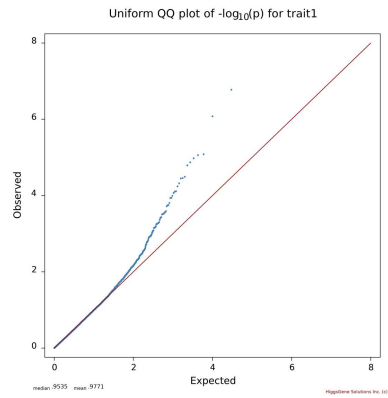
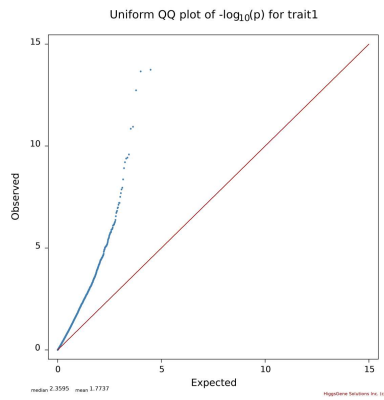
Distribution of $-\log_{10}(p)$ for trait1



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Population Stratification & Structure



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Multiple Testing Issue



H_0 = SNP is not associated with the phenotype ($\alpha = 5\%$)

Probability of type I error for single test is α

Type I error: Incorrect rejection of H_0 (**False positive**)

If there are 1000 tests and tests are independent, then we expect 50 false positive associations!

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Multiple Testing Issue



Controlling type I error rate:

- Permutation
- Bonferroni correction
- False discovery rate
- Positive false discovery rate

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Multiple Testing Issue



Bonferroni correction:

$$\alpha/n$$

Simple but very conservative

$$\alpha = 0.05$$

$$n = 1000$$

Significant level = 0.00005

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Multiple Testing Issue



False Discovery Rate (FDR):

For controlling FDR at 5% level

- Sort p-values from the smallest to the largest
- Find the first p-value that is larger than $(j / n) * 0.05$, where j is the rank of p-value
- Declare all p-values with rank less than j as significant

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Multiple Testing Issue



Positive False Discovery Rate (pFDR)

More complicated than FDR but in some cases better than FDR

With this method p-values are transformed to q-values

<https://www.bioconductor.org/packages/release/bioc/html/qvalue.html>

<https://github.com/StoreyLab/qvalue>

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Ascertainment Bias



- Ascertainment bias is introduced when SNPs are not a random sample of DNA polymorphism
- SNP chips are designed so that well segregating SNP (intermediate allele frequency) are selected (under-representation of rare variants)
- One solution is to impute to the sequence level (not practical yet!)

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GWAS for Sex-Limited Traits



Dairy cattle case:

- Almost all males are genotyped
- Only fraction of females are genotyped
- Calculate EBV for males using daughters information
 - Double counting issue
- Calculate de-regressed EBV for males
 - There could be bias in de-regressed EBV

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Functional Follow-up of GWAS



- In the last decade, a large number of GWAS on high quality phenotypes and big genomic data has resulted in uncovering of numerous SNP association for many traits
- Now, the big challenge is the interpreting the results in biological context

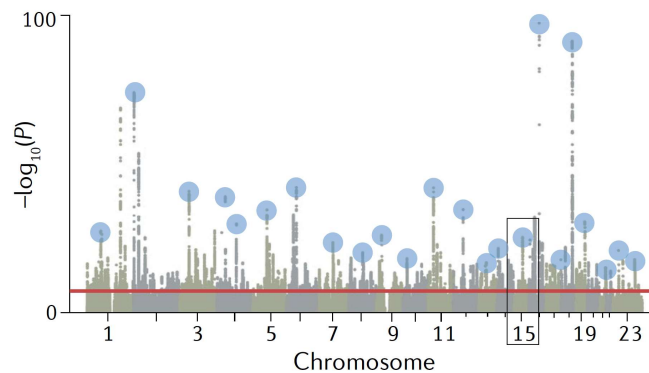
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Functional Follow-up of GWAS



a What are the associated loci?



Uffelmann E., et al., Nature Review (2021) 1:59

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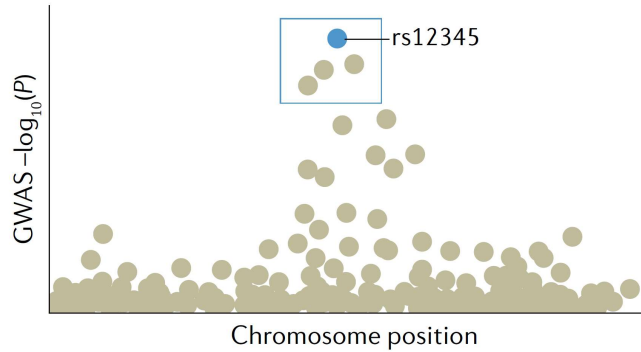
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Functional Follow-up of GWAS



Use statistical fine-mapping to identify the most credible SNP set

What are the likely causal variants?



Uffelmann E., et al., Nature Review (2021) 1:59

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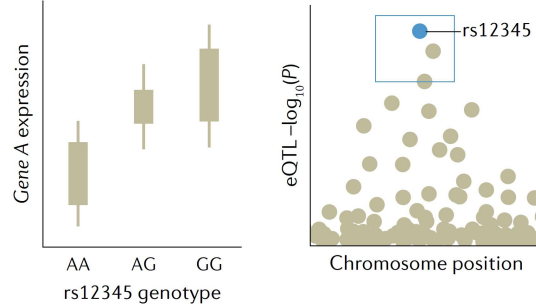
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Functional Follow-up of GWAS



Identify most likely target genes by mapping expression Quantitative Trait Loci (eQTLs)

What are the target genes in the locus?



Uffelmann E., et al., Nature Review (2021) 1:59

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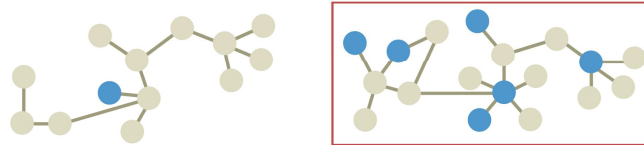
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Functional Follow-up of GWAS



Identify pathways that may mediate the trait using enrichment analysis

What are the affected pathways?



Uffelmann E., et al., Nature Review (2021) 1:59

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Confirmation



Replication in independent samples

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